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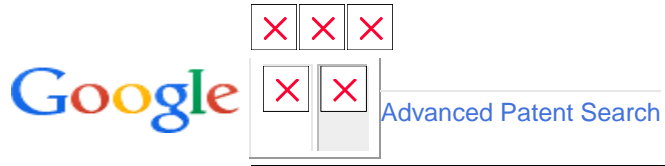
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Patents

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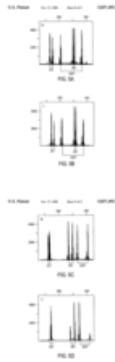
Chromosome 13-linked breast cancer susceptibility gene US 5837492 A

ABSTRACT

The present invention relates generally to the field of human genetics. Specifically, the present invention relates to methods for identifying mutations in the BRCA2 gene and their use in the diagnosis of predisposition to breast cancer. The present invention further relates to methods for identifying mutations in the BRCA2 gene and their use in the diagnosis and prognosis of human cancers. The invention also relates to the therapy of human cancers. The invention also relates to the screening of the BRCA2 gene for mutations, which are useful for diagnosing the predisposition to breast cancer.

IMAGES(9)





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CLAIMS(30)

What is claimed is:

1. An isolated DNA molecule coding for a BRCA2 polypeptide, said DNA molecule comprising a nucleic acid sequence encoded by SEQ ID NO:1.
2. The isolated DNA molecule of claim 1, wherein said DNA molecule comprises the nucleotide sequence set forth in SEQ ID NO:1.
3. The isolated DNA molecule of claim 1, wherein said DNA molecule is an allelic variant of the nucleotide sequence set forth in SEQ ID NO:1.
4. The isolated DNA molecule of claim 1, which contains BRCA2 regulatory sequences.
5. An isolated DNA molecule comprising at least 15 contiguous nucleotides of the DNA molecule of claim 1.
6. An isolated DNA molecule coding for a mutated form of the BRCA2 polypeptide set forth in SEQ ID NO:2, wherein said DNA molecule comprises a nucleic acid sequence encoded by SEQ ID NO:2.
7. The isolated DNA molecule of claim 6, wherein the DNA molecule comprises a mutated nucleotide sequence set forth in SEQ ID NO:2.
8. The isolated DNA molecule of claim 7, wherein the mutation is selected from the group consisting of a deletion, an insertion, a substitution, and a combination thereof.
9. An isolated DNA molecule comprising at least 15 contiguous nucleotides of the DNA of claim 6.
10. The isolated DNA molecule of claim 6 selected from the group consisting of:
 - (a) SEQ ID NO:1 having AC at nucleotide positions 277 and 278 deleted;
 - (b) SEQ ID NO:1 having four nucleotides at positions 982-985 deleted;

- (c) SEQ ID NO:1 having four nucleotides at positions 4706-4709 deleted;
- (d) SEQ ID NO:1 having C at nucleotide position 8525 deleted;
- (e) SEQ ID NO:1 having five nucleotides at positions 9254-9258 deleted;
- (f) SEQ ID NO:1 having GT at nucleotide positions 4075 and 4076 deleted;
- (g) SEQ ID NO:1 having five nucleotides at positions 999-1003 deleted;
- (h) SEQ ID NO:1 having T at nucleotide position 6174 deleted;
- (i) SEQ ID NO:1 having three nucleotides at positions 4132-4134 deleted;
- (j) SEQ ID NO:1 having a C instead of a G at position 451;
- (k) SEQ ID NO:1 having a C instead of an A at position 1093;
- (l) SEQ ID NO:1 having a C instead of a G at position 1291;
- (m) SEQ ID NO:1 having A at position 1493 deleted;
- (n) SEQ ID NO:1 having a T instead of a C at position 2117;
- (o) SEQ ID NO:1 having a C instead of an A at position 2411;
- (p) SEQ ID NO:1 having an A instead of a G at position 4813;
- (q) SEQ ID NO:1 having a G instead of a T at position 5868;
- (r) SEQ ID NO:1 having a T instead of a C at position 5972;
- (s) SEQ ID NO:1 having a T instead of a C at position 6328;
- (t) SEQ ID NO:1 having a T instead of a G at position 7049;
- (u) SEQ ID NO:1 having a C instead of a G at position 7491;
- (v) SEQ ID NO:1 having a G instead of an A at position 9537;
- (w) SEQ ID NO:1 having a T instead of an A at position 10204;
- (x) SEQ ID NO:1 having a G instead of a C at position 10298;

- (y) SEQ ID NO:1 having a G instead of an A at position 10462;
- (z) SEQ ID NO:1 having an A instead of a G at position 203;
- (aa) SEQ ID NO:1 having an A instead of a C at position 1342;
- (bb) SEQ ID NO:1 having a C instead of a T at position 2457;
- (cc) SEQ ID NO:1 having a G instead of an A at position 3199;
- (dd) SEQ ID NO:1 having a G instead of an A at position 3624;
- (ee) SEQ ID NO:1 having a G instead of an A at position 3668;
- (ff) SEQ ID NO:1 having a C instead of a T at position 4035;
- (gg) SEQ ID NO:1 having a G instead of an A at position 7470;
- (hh) SEQ ID NO:1 having a G instead of an A at position 1593;
- (ii) SEQ ID NO:1 having an A instead of a G at position 4296;
- (jj) SEQ ID NO:1 having a G instead of an A at position 5691;
- (kk) SEQ ID NO:1 having a G instead of an A at position 6051;
- (ll) SEQ ID NO:1 having a C instead of a T at position 6828; and
- (mm) SEQ ID NO:1 having a C instead of a T at position 6921.

11. A replicative cloning vector which comprises the isolated DNA molecule of claim 1, or at least 15 contiguous

12. A replicative cloning vector which comprises the isolated DNA molecule of claim 2, or at least 15 contiguous

13. A replicative cloning vector which comprises the isolated DNA molecule of claim 3, or at least 15 contiguous

14. A replicative cloning vector which comprises the isolated DNA molecule of claim 6, or at least 15 contiguous

15. A replicative cloning vector which comprises the isolated DNA molecule of claim 7, or at least 15 contiguous

16. An expression vector which comprises the isolated DNA of claim 1, or at least 15 contiguous nucleotides of

17. An expression vector which comprises the isolated DNA of claim 2, or at least 15 contiguous nucleotides of

18. An expression vector which comprises the isolated DNA of claim 3, or at least 15 contiguous nucleotides of
19. An expression vector which comprises the isolated DNA of claim 6, or at least 15 contiguous nucleotides of
20. An expression vector which comprises the isolated DNA of claim 7, or at least 15 contiguous nucleotides of
21. An isolated host cell transformed with the expression vector of claim 16.
22. An isolated host cell transformed with the expression vector of claim 17.
23. An isolated host cell transformed with the expression vector of claim 18.
24. An isolated host cell transformed with the expression vector of claim 19.
25. An isolated host cell transformed with the expression vector of claim 20.
26. A method of producing recombinant BRCA2 polypeptide which comprises culturing the cells of claim 21 und
27. A method of producing recombinant BRCA2 polypeptide which comprises culturing the cells of claim 22 und
28. A method of producing recombinant BRCA2 polypeptide which comprises culturing the cells of claim 23 und
29. A pair of single-stranded DNA primers of at least 15 nucleotides in length for determination of the nucleotide sequence or at least 15 contiguous nucleotides of the BRCA2 gene.
30. The pair of primers of claim 29 wherein said BRCA2 gene has the nucleotide sequence set forth in SEQ ID N

DESCRIPTION

CROSS REFERENCE TO RELATED APPLICATION

This application is a continuation-in-part of application Ser. No. 08/585,391, filed on 11 Jan. 1996, now abandoned, which is a continuation of application Ser. No. 08/573,779, filed on 18 Dec. 1995, now abandoned, all of which are incorporated herein by reference.

FIELD OF THE INVENTION

The present invention relates generally to the field of human genetics. Specifically, the present invention relates to methods for identifying mutations in the BRCA2 gene and their use in the diagnosis of predisposition to breast cancer. The present invention also relates to the use of the BRCA2 gene in other human cancers and their use in the diagnosis and prognosis of human cancers. The invention also relates to the screening of the BRCA2 gene for mutations, which are useful for diagnosing the predisposition to breast cancer.

The publications and other materials used herein to illuminate the background of the invention, and in particular, cases to p

BACKGROUND OF THE INVENTION

The genetics of cancer is complicated, involving multiple dominant, positive regulators of the transformed state (oncogenes) and multiple negative regulators (tumor suppressor genes). The number of mutations increase beyond fifty (Knudson, 1993).

The involvement of so many genes underscores the complexity of the growth control mechanisms that operate in cells to maintain normal growth. The most common mutations are in the H-ras gene, found in 10-15% of all solid tumors (Anderson et al., 1992). The most frequently mutated gene is p53, which is mutated in 50% of all transformed cells, the dream of a "magic bullet" that can destroy or revert cancer cells while leaving normal tissue unharmed.

The tumor suppressor genes which have been cloned and characterized influence susceptibility to: 1) Retinoblastoma (RB1), 2) Neurofibromatosis type 2A (NF2), 3) Multiple endocrine neoplasia type 2A (MEN2A); and 4) Melanoma (CDKN2).

Tumor suppressor loci that have been mapped genetically but not yet isolated include genes for: Multiple endocrine neoplasia type 2B (MEN2B), Neurofibromatosis type 1 (NF1), Tuberous sclerosis 2 (TSC2). The tumor suppressor genes that have been characterized to date encode products with similarities to cell cycle regulators (CDKN2) and others with no obvious similarity to known proteins (APC and VHL).

In many cases, the tumor suppressor gene originally identified through genetic studies has been shown to be lost or mutated in sporadic cases.

One of the hallmarks of several tumor suppressor genes characterized to date is that they are deleted at high frequency in sporadic cases, either because of a preexisting inherited mutation, or because of a secondary sporadic mutation.

Breast cancer is one of the most significant diseases that affects women. At the current rate, American women have a 1 in 8 chance of developing breast cancer. Ovarian cancer, although less frequent than breast cancer, is often rapidly fatal and is the fourth most common cause of cancer death in women. Breast cancer has been subdivided into two types, early-age onset and late-age onset, based on an inflection in the age-specific incidence rate.

The BRCA1 gene has been isolated (Futreal et al., 1994; Miki et al., 1994) following an intense effort following its mapping to chromosome 17q21.3. BRCA1 confers a high risk of breast cancer but confers a lower risk of ovarian cancer. The remaining susceptibility to early-onset breast cancer is divided between autosomal recessive and autosomal dominant inheritance. BRCA2 is a high risk for breast cancer (Swift et al., 1976; Swift et al., 1991). Late-age onset breast cancer is also often familial although the inheritance pattern is unclear.

Breast cancer has long been recognized to be, in part, a familial disease (Anderson, 1972). Numerous investigators have identified families with a high frequency of breast cancer (Bishop et al., 1988; Newman et al., 1988; Claus et al., 1991). Recent results demonstrate that at least three loci exist which are responsible for the unmapped residual. Hall et al. (1990) indicated that the inherited breast cancer susceptibility in kindreds with a high frequency of breast cancer is due to a single locus.

Most strategies for cloning the chromosome 13-linked breast cancer predisposing gene (BRCA2) require precise genetic localization of the gene. BRCA2 is a high risk of breast cancer. However, cells that contain one wild type BRCA2 allele and one predisposing allele may occasionally suffer loss of heterozygosity. In a two-hit model, predisposing alleles of BRCA2 are recessive, yet susceptibility to cancer is inherited in a dominant fashion: women with one wild type BRCA2 allele and one predisposing allele are not at high risk of cancer. In a one-hit model, predisposing alleles of BRCA2 are dominant, yet susceptibility to cancer is inherited in a recessive fashion: women with two predisposing alleles are at high risk of cancer. In a one-hit model, predisposing alleles of BRCA2 are dominant, yet susceptibility to cancer is inherited in a dominant fashion: women with one predisposing allele are at high risk of cancer. In a two-hit model, predisposing alleles of BRCA2 are recessive, yet susceptibility to cancer is inherited in a dominant fashion: women with one predisposing allele are at high risk of cancer. In a one-hit model, predisposing alleles of BRCA2 are dominant, yet susceptibility to cancer is inherited in a dominant fashion: women with one predisposing allele are at high risk of cancer. In a two-hit model, predisposing alleles of BRCA2 are recessive, yet susceptibility to cancer is inherited in a dominant fashion: women with one predisposing allele are at high risk of cancer.

A second possibility is that BRCA2 predisposing alleles are truly dominant; that is, a wild type allele of BRCA2 cannot overcome the effect of a predisposing allele. In this case, predisposed individuals would undergo some other stochastic change(s) leading to cancer.

The present invention provides an isolated polynucleotide comprising all, or a portion of the BRCA2 locus or of a mutated BRCA2 locus, or a polynucleotide, for example, a recombinant construct suitable for expression in a transformed host cell.

Also provided by the present invention are methods of detecting a polynucleotide comprising a portion of the BRCA2 locus or a mutated BRCA2 locus, or said portion of the BRCA2 locus. The method is useful for either diagnosis of the predisposition to cancer or the diagnosis of cancer.

The present invention also provides isolated antibodies, preferably monoclonal antibodies, which specifically bind to an isolated polynucleotide comprising a portion of the BRCA2 locus or a mutated BRCA2 locus, or said portion of the BRCA2 locus.

The present invention also provides kits for detecting in an analyte a polynucleotide comprising a portion of the BRCA2 locus or a mutated BRCA2 locus, or said portion of the BRCA2 locus.

The present invention further provides methods of preparing a polynucleotide comprising polymerizing nucleotides to yield a polynucleotide comprising a portion of the BRCA2 locus.

The present invention further provides methods of screening the BRCA2 gene to identify mutations. Such methods may further include identifying mutations for use in either diagnosis of the predisposition to cancer or the diagnosis or prognosis of cancer.

The present invention further provides methods of screening suspected BRCA2 mutant alleles to identify mutations in the BRCA2 gene.

In addition, the present invention provides methods of screening drugs for cancer therapy to identify suitable drugs for rest.

Finally, the present invention provides the means necessary for production of gene-based therapies directed at cancer cells. The genes may be reconstituted. Therapeutic agents may also take the form of polypeptides based on either a portion of, or the entire protein.

It is a discovery of the present invention that the BRCA2 locus which predisposes individuals to breast cancer, is a gene encoding a protein. It is a discovery of the present invention that somatic mutations in the BRCA2 locus are also associated with breast cancer, including mutations in the non-coding sequence.

Starting from a region on human chromosome 13 of the human genome, which has a size estimated at about 6 million base pairs, the present invention has identified the BRCA2 locus.

The region containing the BRCA2 locus was identified using a variety of genetic techniques. Genetic mapping techniques including linkage analysis and physical mapping techniques identified the BRCA2 gene. A region which contains the BRCA2 locus is physically bounded by the markers D13S289 and D13S267.

The use of the genetic markers provided by this invention allowed the identification of clones which cover the region from approximately 100 kb to 1.5 Mb. The construction of a contig from a subset of the clones. These P1s, YACs and BACs provide the basis for cloning the BRCA2 gene. The isolation was done using software trapping (a computational method for identifying sequences likely to contain coding sequences). The software trapping was used to obtain sequences of loci expressed in breast and other tissue. These candidate loci were analyzed to identify sequences which are expressed in breast and other tissue. The present invention not only facilitates the early detection of certain cancers, so vital to patient survival, but also permits

Population Resources

Large, well-documented Utah kindreds are especially important in providing good resources for human genetic studies. Each kindred is large enough to confirm the presence of a susceptibility allele. Large sibships are especially important for studying individuals by inference from the haplotypes of their close relatives.

While other populations may also provide beneficial information, such studies generally require much greater effort, and the

Utah's age-adjusted breast cancer incidence is 20% lower than the average U.S. rate. The lower incidence in Utah is probably

Genetic Mapping

Given a set of informative families, genetic markers are essential for linking a disease to a region of a chromosome. Such markers are often polymorphisms based on short tandem repeats (STRs), especially repeats of CpA (Weber and May, 1989; Litt et al., 1989).

Genetic markers useful in searching for a genetic locus associated with a disease can be selected on an ad hoc basis, by choosing markers to determine the ideal distance between genetic markers of a given degree of polymorphism, then selecting markers from libraries which are detected by amplification of the target nucleic acid sequence using PCR; such markers are highly informative, especially

Once linkage has been established, one needs to find markers that flank the disease locus, i.e., one or more markers proximal and distal, as shown in the Examples.

Genetic mapping is usually an iterative process. In the present invention, it began by defining flanking genetic markers around the BRCA2 locus to localize the BRCA2 locus as either distal or proximal to a specific genetic marker (Wooster et al., 1994).

The region surrounding BRCA2, until the disclosure of the present invention, was not well mapped and there were few markers which were polymorphic and which mapped to the BRCA2 region.

Physical Mapping

Three distinct methods were employed to physically map the region. The first was the use of yeast artificial chromosomes

Yeast Artificial Chromosomes (YACs). Once a sufficiently small region containing the BRCA2 locus was identified, physical maps were distributed and contain approximately 50,000 YACs each. The YACs isolated were from these publicly accessible libraries and we selected since they would not have known which YACs were within, and which YACs outside of, the smallest region containing

P1 and BAC Clones. In the present invention, it is advantageous to proceed by obtaining P1 and BAC clones to cover this region, with which the DNA of interest can be manipulated, and improves the signal-to-noise ratio of hybridization assays.

P1 and BAC clones are obtained by screening libraries constructed from the total human genome with specific sequence tags

These P1 and BAC clones can be compared by interspersed repetitive sequence (IRS) PCR and/or restriction enzyme digestion

overlapping contiguous set of clones which covers the region but is not excessively redundant, referred to herein as a "minimal"

P1 clones (Stenberg, 1990; Sternberg et al., 1990; Pierce et al., 1992; Shizuya et al., 1992) were isolated by Genome Sciences. This method permitted the covering of the genomic region with an independent set of clones not derived from YACs. This guards against

Gene Isolation.

There are many techniques for testing genomic clones for the presence of sequences likely to be candidates for the coding

(a) Zoo blots. The first technique is to hybridize cosmids to Southern blots to identify DNA sequences which are evolutionarily conserved. A variety of species are commercially available (Clontech, Cat. 7753-1).

(b) Identifying HTF islands. The second technique involves finding regions rich in the nucleotides C and G, which often occur

(c) Exon trapping. The third technique is exon trapping, a method that identifies sequences in genomic DNA which contain exons. The selection of RNA sequences which are flanked by functional 5' and/or 3' splice sites. The products of the exon amplification are then screened for exon trapping.

(d) Hybridizing cDNA to P1s, BACs or YACs. The fourth technique is a modification of the selective enrichment technique. Modified for the present purpose, involves binding DNA from the region of BRCA2 present in a YAC to a column matrix and then hybridizing with cDNA.

(e) Identification of cDNAs. The fifth technique is to identify cDNAs that correspond to the BRCA2 locus. Hybridization probes

Another variation on the theme of direct selection of cDNA can be used to find candidate genes for BRCA2 (Lovett et al., 1993). This involves binding sites for primers in subsequent PCR amplification reactions using biotinylated primers. Target cDNA is generated from a library of cDNAs. These adapters serve as amplification sites for PCR. The target and probe sequences are denatured and mixed with PCR reagents. High stringency and the retained cDNAs are eluted and amplified by PCR. The selected cDNA is subjected to further rounds of selection.

Testing the cDNA for Candidacy

Proof that the cDNA is the BRCA2 locus is obtained by finding sequences in DNA extracted from affected kindred members and comparing them to non-kindred individuals with breast cancer then in individuals in the general population. Finally, since tumors often mutate BRCA2 sequences in tumor tissue. Whether one is comparing BRCA2 sequences from tumor tissue to BRCA2 alleles from the germline of the same individual or to BRCA2 alleles from other individuals. These mutations can take a number of forms. The most severe forms would be frame shift mutations or large deletions which would have a significant effect on the protein produced, such as changes to or from a cysteine residue, from a basic amino acid to an acidic amino acid. Conservative amino acid substitutions would not generally be expected to disrupt protein function.

According to the diagnostic and prognostic method of the present invention, alteration of the wild-type BRCA2 locus is detected by comparing BRCA2 sequences from tumor tissue to BRCA2 alleles from the germline of the same individual or to BRCA2 alleles from other individuals. including deletions, insertions and point mutations in the coding and noncoding regions. Deletions may be of the entire gene or of a portion of the gene.

inherited in the germline. Germline mutations can be found in any of a body's tissues and are inherited. If only a single allele information. A BRCA2 allele which is not deleted (e.g., found on the sister chromosome to a chromosome carrying a BRCA2 product. However, mutations leading to non-functional gene products would also lead to a cancerous state. Point mutations in BRCA2 gene product, or to a decrease in mRNA stability or translation efficiency.

Useful diagnostic techniques include, but are not limited to fluorescent in situ hybridization (FISH), direct DNA sequencing,

Predisposition to cancers, such as breast cancer, and the other cancers identified herein, can be ascertained by testing an individual's body. Most simply, blood can be drawn and DNA extracted from the cells of the blood. In addition, prenatal diagnosis can be discussed herein.

There are several methods that can be used to detect DNA sequence variation. Direct DNA sequencing, either manual sequencing, is rarely missed. Another approach is the single-stranded conformation polymorphism assay (SSCA) (Orita et al., 1989). This method has increased throughput possible with SSCA makes it an attractive, viable alternative to direct sequencing for mutation detection. Other methods which affect two complementary DNA strands include clamped denaturing gel electrophoresis (CDGE) (Sheffield et al., 1991), heteroduplex analysis, which affects transcription or translation of the protein. Other methods which might detect these classes of mutations such as Southern blotting, a recent review by Grompe (1993). Once a mutation is known, an allele specific detection approach such as allele specific oligonucleotide (ASO) analysis.

In order to detect the alteration of the wild-type BRCA2 gene in a tissue, it is helpful to isolate the tissue free from surrounding normal cells by cytometry. These techniques, as well as other techniques for separating tumor cells from normal cells, are well known in the art.

A rapid preliminary analysis to detect polymorphisms in DNA sequences can be performed by looking at a series of Southern blots of DNA hybridizing fragments (differing in length from control DNA when probed with sequences near or including the BRCA2 locus).

Detection of point mutations may be accomplished by molecular cloning of the BRCA2 allele(s) and sequencing the allele(s). The presence of a mutation then be determined.

There are six well known methods for a more complete, yet still indirect, test for confirming the presence of a susceptibility gene mutation: 1) Southern blotting (Orita et al., 1991); 2) allele-specific oligonucleotides (ASOs) (Conner et al., 1983); 3) the use of proteins which recognize nucleotide mismatches; 4) BRCA2 mutation is not present, an amplification product is not observed. Amplification Refractory Mutation System (ARMS) and 5) restriction fragment length polymorphism (RFLP) probes for the gene or surrounding marker genes can be used to score a mutation.

Such a method is particularly useful for screening relatives of an affected individual for the presence of the BRCA2 mutation.

In the first three methods (SSCA, DGGE and RNase protection assay), a new electrophoretic band appears. SSCA detects differences in migration rates of mutant sequences compared to wild-type sequences, using a denaturing gradient gel electrophoresis (DGGE) method. DGGE detects differences in migration rates of mutant sequences compared to wild-type sequences, using a denaturing gradient gel electrophoresis (DGGE) method. A protein binds only to sequences that contain a nucleotide mismatch in a heteroduplex between mutant and wild-type sequences.

Mismatches according to the present invention, are hybridized nucleic acid duplexes in which the two strands are not 100% complementary. Because, more sensitive than sequencing, they are simpler to perform on a large number of tumor samples. An example of a mismatch cleavage site is shown in Figure 1.

and either mRNA or DNA isolated from the tumor tissue are annealed (hybridized) together and subsequently digested with an endonuclease. In an electrophoretic gel matrix, if a mismatch has been detected and cleaved by RNase A, an RNA product will be seen which is complementary to the BRCA2 mRNA or gene, it will be desirable to use a number of these probes to screen the whole mRNA sequence for mismatches.

In similar fashion, DNA probes can be used to detect mismatches, through enzymatic or chemical cleavage. See, e.g., Cottrell et al. (1990). Using riboprobes or DNA probes, the cellular mRNA or DNA which might contain a mutation can be amplified using PCR (see below).

DNA sequences of the BRCA2 gene which have been amplified by use of PCR may also be screened using allele-specific probes. By use of a battery of such allele-specific probes, PCR amplification products can be screened. A positive result with a probe under stringent hybridization conditions indicates the presence of the same mutation in the tumor tissue as in the allele.

The most definitive test for mutations in a candidate locus is to directly compare genomic BRCA2 sequences from cancer patients to those from control individuals.

Mutations from cancer patients falling outside the coding region of BRCA2 can be detected by examining the non-coding region. An abnormal size or abundance in cancer patients as compared to control individuals.

Alteration of BRCA2 mRNA expression can be detected by any techniques known in the art. These include Northern blot analysis and Western blot analysis of BRCA2 protein. For example, monoclonal antibodies immunoreactive with BRCA2 can be used to screen a tissue. Lack of immunoreactivity in the art. These include Western blots, immunohistochemical assays and ELISA assays. Any means for detecting an alteration in BRCA2 mRNA or a mutant BRCA2 gene product indicates alteration of a wild-type BRCA2 gene.

Mutant BRCA2 genes or gene products can also be detected in other human body samples, such as serum, stool, urine and sweat. In addition, the BRCA2 gene product itself may be secreted into the extracellular space and found in these body samples even in the absence of such body samples for mutant BRCA2 genes or gene products.

The methods of diagnosis of the present invention are applicable to any tumor in which BRCA2 has a role in tumorigenesis.

The primer pairs of the present invention are useful for determination of the nucleotide sequence of a particular BRCA2 allele. By use of these primers allows synthesis of all of the nucleotides of the BRCA2 gene coding sequences, i.e., the exons. The set of primers can be used to identify a mutant allele as a template.

In order to facilitate subsequent cloning of amplified sequences, primers may have restriction enzyme site sequences appended to them. These are well known in the art. The primers themselves can be synthesized using techniques which are well known in the art. General methods for synthesis, in addition to those disclosed below, is well within the skill of the art.

The nucleic acid probes provided by the present invention are useful for a number of purposes. They can be used in Southern blot analysis with the BRCA2 gene or mRNA using other techniques.

It has been discovered that individuals with the wild-type BRCA2 gene do not have cancer which results from the BRCA2 gene mutation or altered function, directly correlates to an increased risk of cancer. In order to detect a BRCA2 gene mutation, a biological

described above. The mutant alleles are then sequenced to identify the specific mutation of the particular mutant allele. All mutations, especially those which lead to an altered function of the BRCA2 protein, are then used for the diagnostic and prognostic purposes.

Definitions

The present invention employs the following definitions:

"Amplification of Polynucleotides" utilizes methods such as the polymerase chain reaction (PCR), ligation amplification (or PCR); and Wu et al., 1989a (for LCR). Reagents and hardware for conducting PCR are commercially available. Primers used for amplification may be sequenced directly. Alternatively, but less desirably, the amplified sequence(s) may be cloned prior to sequencing.

"Analyte polynucleotide" and "analyte strand" refer to a single- or double-stranded polynucleotide which is suspected of containing a mutation.

"Antibodies." The present invention also provides polyclonal and/or monoclonal antibodies and fragments thereof, and immunoassays thereof. The term "antibody" is used both to refer to a homogeneous molecular entity, or a mixture such as a serum produced in rabbits. Rabbit sera is tested for immunoreactivity to the BRCA2 polypeptide or fragment. Monoclonal antibodies may be made by Harlow & Lane, 1988. These antibodies will be useful in assays as well as pharmaceuticals.

Once a sufficient quantity of desired polypeptide has been obtained, it may be used for various purposes. A typical use is to select an appropriate target immune system, typically mouse or rabbit, is selected. Substantially purified antigen is presented to the immune system. In some cases, other species may be substituted for mouse or rabbit. Polyclonal antibodies are then purified using techniques known in the art.

An immunological response is usually assayed with an immunoassay. Normally, such immunoassays involve some purification of the antigen.

Monoclonal antibodies with affinities of $10^{-8} M^{-1}$ or preferably 10^{-9} to $10^{-1} M^{-1}$ or stronger will typically be made by standard techniques. Spleen cells excised and individual spleen cells fused, typically, to immortalized myeloma cells under appropriate selection conditions. The resulting hybridomas are then screened for the desired antibody.

Other suitable techniques involve in vitro exposure of lymphocytes to the antigenic polypeptides, or alternatively, to selected antigens, and subsequent joining, either covalently or non-covalently, a substance which provides for a detectable signal. A wide variety of labels and detection systems, including magnetic particles and the like. Patents teaching the use of such labels include U.S. Pat. Nos. 3,817,837; 3,850,752; 3,930,451.

"Binding partner" refers to a molecule capable of binding a ligand molecule with high specificity, as for example, an antigen-antibody hybridization) under the isolation conditions. Specific binding partners are known in the art and include, for example, biotin-antibiotin complexes, about 15 bases in length, and may be at least 40 bases in length. The polynucleotides may be composed of DNA, RNA, or a hybrid of DNA and RNA.

A "biological sample" refers to a sample of tissue or fluid suspected of containing an analyte polynucleotide or polypeptide or in vitro cell culture constituents.

As used herein, the terms "diagnosing" or "prognosing," as used in the context of neoplasia, are used to indicate 1) the classification of a patient as having or not having a particular disease, and 2) the prediction of the course of a disease.

"Encode". A polynucleotide is said to "encode" a polypeptide if, in its native state or when manipulated by methods well known in the art, the amino acid sequence of the polypeptide can be deduced therefrom.

"Isolated" or "substantially pure". An "isolated" or "substantially pure" nucleic acid (e.g., an RNA, DNA or a mixed polymer) embraces a nucleic acid sequence or protein which has been removed from its naturally occurring environment, and includes

"BRCA2 Allele" refers to normal alleles of the BRCA2 locus as well as alleles carrying variations that predispose individuals

"BRCA2 Locus," "BRCA2 Gene," "BRCA2 Nucleic Acids" or "BRCA2 Polynucleotide" each refer to polynucleotides, all of which are capable of initiating and/or progression of other types of tumors. The locus is indicated in part by mutations that predispose individuals to cancer. The BRCA2 locus is intended to include all allelic variations of the DNA sequence.

These terms, when applied to a nucleic acid, refer to a nucleic acid which encodes a BRCA2 polypeptide, fragment, homologue, or has substantial homology with a natural BRCA2-encoding gene or a portion thereof. The coding sequence for a BRCA2 polypeptide

The polynucleotide compositions of this invention include RNA, cDNA, genomic DNA, synthetic forms, and mixed polymers. Examples include, for example, labels, methylation, substitution of one or more of the naturally occurring nucleotides with an analog, internucleotide linkages (e.g., phosphorothioates, phosphoramidates, phosphonates, etc.), polypeptides), intercalators (e.g., acridine, psoralen, etc.), chelators, alkylators, and modified linkages (e.g., alpha anomeric linkages). Examples include, for example, those in which peptide linkages substitute for phosphate linkages in the backbone of the molecule.

The present invention provides recombinant nucleic acids comprising all or part of the BRCA2 region. The recombinant nucleic acid is a polynucleotide of genomic, cDNA, semi-synthetic, or synthetic origin which, by virtue of its origin or manipulation, 1) is not a naturally occurring

Therefore, recombinant nucleic acids comprising sequences otherwise not naturally occurring are provided by this invention.

cDNA or genomic libraries of various types may be screened as natural sources of the nucleic acids of the present invention. Phage libraries are normally preferred, but other types of libraries may be used. Clones of a library

The DNA sequences used in this invention will usually comprise at least about five codons (nucleotides), more usually at least about ten, and specifically with a BRCA2-encoding sequence.

Techniques for nucleic acid manipulation are described generally, for example, in Sambrook et al., 1989 or Ausubel et al., 1989, Promega Biotec, U. S. Biochemicals, New England Nuclear, and a number of other sources. The recombinant nucleic acids are used as probes. See, GenBank, National Institutes of Health.

"BRCA2 Region" refers to a portion of human chromosome 13 bounded by the markers tdj3820 and YS-G-B10T. This region

As used herein, the terms "BRCA2 locus," "BRCA2 allele" and "BRCA2 region" all refer to the double-stranded DNA component

As used herein, a "portion" of the BRCA2 locus or region or allele is defined as having a minimal size of at least about eight

"BRCA2 protein" or "BRCA2 polypeptide" refer to a protein or polypeptide encoded by the BRCA2 locus, variants or fragments thereof. This term also does not refer to, or exclude modifications of the polypeptide, for example, glycosylations, acetylations, phosphorylations, and other modifications known in the art, both naturally and non-naturally occurring. Ordinarily, such polypeptides will be at least 16 amino acids long. Under stringent conditions, to BRCA2-encoding nucleic acids and closely related polypeptides or proteins retrieved by antisera to BRCA2 protein.

The length of polypeptide sequences compared for homology will generally be at least about 16 amino acids, usually at least about 20 amino acids.

"Operably linked" refers to a juxtaposition wherein the components so described are in a relationship permitting them to function together in the performance of the desired function.

"Probes" . Polynucleotide polymorphisms associated with BRCA2 alleles which predispose to certain cancers or are associated with certain cancers. In order that the probes will be perfectly complementary to the target sequence, stringent conditions will be used. Hybridization stringency conditions are those which minimize noise. Since such indications identify neutral DNA polymorphisms as well as mutations, these indications are not limited to mutations.

Probes for BRCA2 alleles may be derived from the sequences of the BRCA2 region or its cDNAs. The probes may be of any length, but the range of about 8-30 base pairs, since the hybrid will be relatively stable under even stringent conditions. If some degree of stringency is required, the length of the probe should be at least about 8 base pairs.

The probes will include an isolated polynucleotide attached to a label or reporter molecule and may be used to isolate other polynucleotides using homologous or complementary polynucleotides. Alternatively, polynucleotides encoding these or similar polypeptides may be synthesized. Mutations may be introduced to modify the properties of the polypeptide, perhaps to change ligand-binding affinities, interactions with other molecules, or other properties.

Probes comprising synthetic oligonucleotides or other polynucleotides of the present invention may be derived from natural or synthetic sources.

Portions of the polynucleotide sequence having at least about eight nucleotides, usually at least about 15 nucleotides, and up to the full length of the polynucleotide sequence may be used. The polynucleotide may be in a purified form or in a tissue.

"Protein modifications or fragments" are provided by the present invention for BRCA2 polypeptides or fragments thereof which are modified by acetylation, carboxylation, phosphorylation, glycosylation, ubiquitination, labeling, e.g., with radionuclides, and various enzymatic modifications.

A variety of methods for labeling polypeptides and of substituents or labels useful for such purposes are well known in the art. The labeled ligand. The choice of label depends on the sensitivity required, ease of conjugation with the primer, stability requirements, and other factors.

Besides substantially full-length polypeptides, the present invention provides for biologically active fragments of the polypeptide system, as well as sharing of immunological epitopes for binding, serving as either a competitor or substitute antigen for an antibody. Such fragments consists of at least five such amino acids, and more usually consists of at least 8-10 such amino acids. Methods of determining the biological activity of such fragments are well known in the art.

For immunological purposes, tandem-repeat polypeptide segments may be used as immunogens, thereby producing highly immunogenic polypeptides.

The present invention also provides for fusion polypeptides, comprising BRCA2 polypeptides and fragments thereof. Homologous domains or properties or activities of the derivative proteins. For example, ligand-binding or other domains may be "swapped" between polypeptides, e.g., galactosidase, trpE, protein A, β -lactamase, alpha amylase, alcohol dehydrogenase and yeast alpha mating factor. See, e.g., U.S. Pat. No. 4,889,412.

to various substitutions, deletions and other modifications. Conservative substitutions typically include substitutions within t

A polypeptide "fragment," "portion" or "segment" is a stretch of amino acid residues of at least about five to seven contiguous

The polypeptides of the present invention, if soluble, may be coupled to a solid-phase support, e.g., nitrocellulose, nylon, c

"Target region" refers to a region of the nucleic acid which is amplified and/or detected. The term "target sequence" refers t

The practice of the present invention employs, unless otherwise indicated, conventional techniques of chemistry, molecular biology, and materials for human gene mapping, including mapping of human chromosome 13, is provided, e.g., in White and Lalou

Preparation of recombinant or chemically synthesized nucleic acids; vectors, transformation, host cells

Large amounts of the polynucleotides of the present invention may be produced by replication in a suitable host cell. Naturally occurring polynucleotides are usually the polynucleotide constructs will be suitable for replication in a unicellular host, such as yeast or bacteria, but may be described, e.g., in Sambrook et al., 1989 or Ausubel et al., 1992.

The polynucleotides of the present invention may also be produced by chemical synthesis, e.g., by the phosphoramidite method. The polynucleotides obtained from the single-stranded product of chemical synthesis either by synthesizing the complementary strand and annealing

Polynucleotide constructs prepared for introduction into a prokaryotic or eukaryotic host may comprise a replication system, a promoter, an encoding segment. Expression vectors may include, for example, an origin of replication or autonomously replicating sequence, a poly(A) mRNA stabilizing sequences. Secretion signals may also be included where appropriate, whether from a native BRCA2 protein or otherwise. Such vectors may be prepared by means of standard recombinant techniques well known in the art and discussed, for example, in Sambrook et al., 1989.

An appropriate promoter and other necessary vector sequences will be selected so as to be functional in the host, and may be obtained from such vendors as Stratagene, New England Biolabs, etc. Other promoters might include the early and late promoters from SV40 (Fiers et al., 1978) or promoters derived from murine Moloney retroviruses. For appropriate enhancer and other expression control sequences, see also Enhancers and Eukaryotic Gene Expression Vectors, Sambrook et al., 1989.

While such expression vectors may replicate autonomously, they may also replicate by being inserted into the genome of the host cell.

Expression and cloning vectors will likely contain a selectable marker, a gene encoding a protein necessary for survival or growth in the presence of substances, e.g. ampicillin, neomycin, methotrexate, etc.; b) complement auxotrophic deficiencies, or c) supply critical nutrients. See Sambrook et al., 1989, for art.

The vectors containing the nucleic acids of interest can be transcribed in vitro, and the resulting RNA introduced into the host cell by electroporation; transfection employing calcium chloride, rubidium chloride, calcium phosphate, DEAE-dextran, or other suitable methods. The polynucleotides into the host cell by any method known in the art, including, inter alia, those described above, will be referred to as "transformation."

Large quantities of the nucleic acids and polypeptides of the present invention may be prepared by expressing the BRCA2 gene in *Bacillus subtilis* or *Pseudomonas* may also be used.

Mammalian or other eukaryotic host cells, such as those of yeast, filamentous fungi, plant, insect, or amphibian or avian species are VERO and HeLa cells, Chinese hamster ovary (CHO) cells, and WI38, BHK, and COS cell lines, although it will be apparent to those skilled in the art that other cells may be used.

Clones are selected by using markers depending on the mode of the vector construction. The marker may be on the same plasmid as the BRCA2 gene. Temperature sensitivity may also serve as an appropriate marker.

Prokaryotic or eukaryotic cells transformed with the polynucleotides of the present invention will be useful not only for the production of the polypeptides but also for the study of the function of the BRCA2 gene.

Antisense polynucleotide sequences are useful in preventing or diminishing the expression of the BRCA2 locus, as will be apparent to those skilled in the art. An antisense construct with a control of a promoter in an antisense orientation and introduced into a cell. Expression of such an antisense construct will result in the inhibition of the BRCA2 gene.

The probes and primers based on the BRCA2 gene sequences disclosed herein are used to identify homologous BRCA2 genes in other organisms. The probes and primers are also used to identify BRCA2 genes isolated from other organisms.

Methods of Use: Nucleic Acid Diagnosis and Diagnostic Kits

In order to detect the presence of a BRCA2 allele predisposing an individual to cancer, a biological sample such as blood or saliva from the individual is prepared and analyzed for the presence or absence of mutant alleles of BRCA2. Results of these tests are provided to health care providers or to private individuals for self-diagnosis.

Initially, the screening method involves amplification of the relevant BRCA2 sequences. In another preferred embodiment of the present invention, the target sequences can be detected with a high level of sensitivity.

The most popular method used today is target amplification. Here, the target nucleic acid sequence is amplified with polymerase chain reaction (PCR). The target sequence increases in copy number through the use of polymerase-driven amplification cycles. Once amplified, the resulting nucleic acid can be detected with a high level of sensitivity.

When the probes are used to detect the presence of the target sequences (for example, in screening for cancer susceptibility), the target sequences are usually detected by denaturation, restriction digestion, electrophoresis or dot blotting. The targeted region of the analyte nucleic acid usually must be denatured. Denaturation can be carried out by various techniques known in the art.

Analyte nucleic acid and probe are incubated under conditions which promote stable hybrid formation of the target sequence. High stringency conditions are desirable in order to prevent false positives. However, conditions of high stringency are used only when the target sequence is known. Factors such as temperature, ionic strength, base composition, probe length, and concentration of formamide. These factors are outlined in the following section.

Detection, if any, of the resulting hybrid is usually accomplished by the use of labeled probes. Alternatively, the probe may be labeled with a fluorescent label which may be incorporated by known methods (e.g., nick translation, random priming or kinasing), biotin, fluorescent antibody, or other methods known in the art.

hybrids to be detected from extraneous materials and/or that amplify the signal from the labeled moiety. A number of these

As noted above, non-PCR based screening assays are also contemplated in this invention. An exemplary non-PCR based enzyme covalently linked to the probe, such that the covalent linkage does not interfere with the specificity of the hybridization development or luminescent output resulting in a 10^3 - 10^6 increase in sensitivity. For an example relating to preparation of

Two-step label amplification methodologies are known in the art. These assays work on the principle that a small ligand (such as an antibody) specifically recognizes a target. Allele-specific probes are also contemplated within the scope of this example, and exemplary allele specific probes

In one example, the small ligand attached to the nucleic acid probe is specifically recognized by an antibody-enzyme conjugate for labeling nucleic acid probes according to this embodiment see Martin et al., 1990. In a second example, the small ligand is specifically recognized by a protein such as avidin for labeling probes and their use in biotin-avidin based assays see Rigby et al., 1977 and Nguyen et al., 1992.

It is also contemplated within the scope of this invention that the nucleic acid probe assays of this invention will employ a collection of probes is alternatively 2, 3, or 5 different nucleic acid probe sequences. In another example, to detect the presence of mutations or alterations in BRCA2. In this embodiment, any number of probes can be used, and will preferably include probes corresponding to the wild-type BRCA2 and those that have the BRCA2 regions shown in SEQ ID NO:1 and FIG. 3, both 5' and 3' to the mutation site.

Methods of Use: Peptide Diagnosis and Diagnostic Kits

The neoplastic condition of lesions can also be detected on the basis of the alteration of wild-type BRCA2 polypeptide. Such antibodies may be prepared as discussed above under the heading "Antibodies" and as further shown in Examples 9 and 10. Such antibodies may immunoprecipitate BRCA2 proteins from solution as well as react with BRCA2 protein on Western or immunoblots of polyacrylamide gels.

Preferred embodiments relating to methods for detecting BRCA2 or its mutations include enzyme linked immunosorbent assays as described by David et al. in U.S. Pat. Nos. 4,376,110 and 4,486,530, hereby incorporated by reference, and exemplified in Example 9.

Methods of Use: Drug Screening

This invention is particularly useful for screening compounds by using the BRCA2 polypeptide or binding fragment thereof.

The BRCA2 polypeptide or fragment employed in such a test may either be free in solution, affixed to a solid support, or bound to cells. Such assays. Such cells, either in viable or fixed form, can be used for standard binding assays. One may measure, for example, the binding of a labeled agent, which is not interfered with by the agent being tested.

Thus, the present invention provides methods of screening for drugs comprising contacting such an agent with a BRCA2 polypeptide or fragment by methods well known in the art. In such competitive binding assays the BRCA2 polypeptide or fragment is typically labeled with a radioisotope, a fluorescent dye, or a biotin. BRCA2: ligand binding, respectively.

Another technique for drug screening provides high throughput screening for compounds having suitable binding affinity to

substrate, such as plastic pins or some other surface. The peptide test compounds are reacted with BRCA2 polypeptide and the antibodies to the polypeptide can be used to capture antibodies to immobilize the BRCA2 polypeptide on the solid phase.

This invention also contemplates the use of competitive drug screening assays in which neutralizing antibodies capable of binding to more antigenic determinants of the BRCA2 polypeptide.

A further technique for drug screening involves the use of host eukaryotic cell lines or cells (such as described above) which are transfected with the test compounds to determine if the compound is capable of regulating the growth of BRCA2 defective cells.

Methods of Use: Rational Drug Design

The goal of rational drug design is to produce structural analogs of biologically active polypeptides of interest or of small molecules that mimic the polypeptide in vivo. See, e.g., Hodgson, 1991. In one approach, one first determines the three-dimensional structure of a polypeptide. Information regarding the structure of a polypeptide may be gained by modeling based on the structure of homologous proteins. An amino acid residue may be replaced by Ala, and its effect on the peptide's activity is determined. Each of the amino acid residues of the peptide is analyzed in a similar manner.

It is also possible to isolate a target-specific antibody, selected by a functional assay, and then to solve its crystal structure. The structure of a pharmacologically active antibody. As a mirror image of a mirror image, the binding site of the anti-ids would be expected to be a mirror image of the target.

Thus, one may design drugs which have, e.g., improved BRCA2 polypeptide activity or stability or which act as inhibitors, and which are designed using crystallography. In addition, the knowledge of the BRCA2 protein sequence provided herein will guide those employing conventional methods.

Methods of Use: Gene Therapy

According to the present invention, a method is also provided of supplying wild-type BRCA2 function to a cell which carries a mutant BRCA2 gene on an extrachromosomal. In such a situation, the gene will be expressed by the cell from the extrachromosomal location. If a gene which carries the wild-type BRCA2 gene or a part thereof is introduced into the mutant cell in such a way that it recombines with the endogenous BRCA2 gene, extrachromosomal maintenance are known in the art, and any suitable vector may be used. Methods for introducing DNA into cells are known in the art and may be used as model systems to study cancer remission and drug treatments which promote such remission.

As generally discussed above, the BRCA2 gene or fragment, where applicable, may be employed in gene therapy methods for the treatment of cells in which the gene is absent or diminished compared to normal cells. It may also be useful to increase the level of expression of a given BRCA2 gene.

Gene therapy would be carried out according to generally accepted methods, for example, as described by Friedman, 1992. A copy of the BRCA2 gene linked to expression control elements and capable of replicating inside the tumor cells, is prepared and introduced into any tumor cells that may have metastasized to other sites). If the transfected gene is not permanently incorporated into the genome, the gene may be re-introduced into the tumor cells.

Gene transfer systems known in the art may be useful in the practice of the gene therapy methods of the present invention. Such systems include adenoviruses (Kapikian, 1992; Quantin et al., 1992; Rosenfeld et al., 1992; Wilkinson et al., 1992; Stratford-Perricaudet et al., 1990), herpesviruses (Miller, 1992; Miller et al., 1992), and retroviruses of avian (Bradyopadhyay and Temin, 1984; Petropoulos et al., 1992), murine (Miller, 1992; Miller et al., 1992), and human (Miller, 1992; Miller et al., 1992).

been based on disabled murine retroviruses.

Nonviral gene transfer methods known in the art include chemical techniques such as calcium phosphate coprecipitation (Cristofalo et al., 1987) and electroporation (Wang and Hoffman, 1987) as well as delivery via liposomes (Felgner et al., 1987; Wang and Huang, 1989; Kaneda et al., 1989; Stewart et al., 1992; Nabel et al., 1990; Lippman et al., 1991a; Curiel et al., 1991b). Viral-mediated gene transfer can be combined with direct in vivo gene transfer using liposomes. Producer cells would then provide a continuous source of vector particles. This technique has been approved for use in humans.

In an approach which combines biological and physical gene transfer methods, plasmid DNA of any size is combined with liposomes. The liposomes facilitate the internalization, and degradation of the endosome before the coupled DNA is damaged.

Liposome/DNA complexes have been shown to be capable of mediating direct in vivo gene transfer. While in standard liposomes, the DNA is degraded by the acidic environment of the endosome.

Gene transfer techniques which target DNA directly to breast and ovarian tissues, e.g., epithelial cells of the breast or ovarian cells, are known in the art. Such techniques are based on the presence of the corresponding ligand receptors on the cell surface of the target cell/tissue type. One appropriate receptor is the epidermal growth factor receptor. Upon binding and internalization of the DNA-protein complex occurs. To overcome the problem of intracellular destruction of DNA, the DNA is protected by a protein coat.

The therapy involves two steps which can be performed singly or jointly. In the first step, prepubescent females who carry a mutant BRCA2 allele are treated with a BRCA2-producing mammalian cell. In the second step, the treated individuals have reduced risk of breast cancer to the extent that the effect of the susceptible allele has been counteracted. The therapy has been shown to have beneficial effects on the breast of a full term pregnancy.

Methods of Use: Peptide Therapy

Peptides which have BRCA2 activity can be supplied to cells which carry mutant or missing BRCA2 alleles. The sequence of the peptide is chosen to be complementary to the BRCA2-producing mammalian cells. In addition, the techniques of synthetic chemistry can be employed to synthesize BRCA2 in a microorganism or in vitro.

Active BRCA2 molecules can be introduced into cells by microinjection or by use of liposomes, for example. Alternatively, the use of a BRCA2-producing mammalian cell can be used for the reversal of the neoplastic state. Other molecules with BRCA2 activity (for example, peptides, drugs or organic compounds) can be used for the same purpose.

Methods of Use: Transformed Hosts

Similarly, cells and animals which carry a mutant BRCA2 allele can be used as model systems to study and test for substances which can reverse the neoplastic state. Cells which carry the mutation in the BRCA2 allele, as described above. After a test substance is applied to the cells, the neoplastically transformed cells are cultured. Assays for each of these traits are known in the art.

Animals for testing therapeutic agents can be selected after mutagenesis of whole animals or after treatment of germline cells. The mutation can be disrupted by insertion or deletion mutation or other genetic alterations using conventional techniques (Capecchi, 1989; Valasek et al., 1990). The presence of tumors must be assessed. If the test substance prevents or suppresses the growth of tumors, then the test substance is a candidate for further study.

The present invention is described by reference to the following Examples, which are offered by way of illustration and are not intended to limit the scope of the invention.

EXAMPLE 1 Ascertain and Study Kindreds Likely to Have a Chromosome 13-Linked Breast Cancer Susceptibility Locus

Extensive cancer prone kindreds were ascertained from a defined population providing a large set of extended kindreds with opportunity for informative recombinants to occur within the small region being investigated. This vastly improved the chance

Each kindred was extended through all available connecting relatives, and to all informative first degree relatives of each person reported in the kindred which were not confirmed in the Utah Cancer Registry were researched. Medical records or death certificates and relatives of deceased cases so that the genotype of the deceased cases could be inferred from the genotypes of their

Kindreds which had three or more cancer cases with inferable genotypes were selected for linkage studies to chromosome 13. Kindreds with two sisters or a mother and her daughter with breast cancer. Additionally, kindreds which have been studied since 1980 as well as kindreds were investigated and expanded in our clinic in the manner described above.

For each sample collected in these kindreds, DNA was extracted from blood or paraffin- embedded tissue blocks using standard methods. To aid in this effort, STR markers on chromosome 13 were developed by screening a chromosome specific cosmid library and published reports, or as part of the Breast Cancer Linkage Consortium, or from other investigators. All genotyping films were

LOD scores for each kindred were calculated for two recombination fraction values, 0.001 and 0.1. (For calculation of LOD scores, specific risks for breast cancer in non-gene carriers. Allele frequencies for the markers used for the LOD score calculations were

Kindred 107 is the largest chromosome 13-linked breast cancer family reported to date by any group. The evidence of linkage is

In order to improve the characterization of our recombinants and define closer flanking markers, a dense map of this region was constructed showing the location of ten markers used in the genetic analysis. Table 1 gives the LOD scores for linkage for each of the 19

TABLE 1

D13SKindred	FBR	MBR	OV	LOD	Probability (2)	3820	4247	260
267								107* 22 3 2 5.06
82043*	2 1 1	0.86	0.98	6 30 3	12 7 10 5 8 4	122018	3 1 0 n.d.	0.90 9 12 7 3 8 3 6 6 5 8 937
1 0	n.d.	0.90	3 29 7	10 5 8 5 5 5	88003	2 1 0 n.d.	0.90 4 12 6 10 6 3 4 5 4	82367 6 0 1 0.4
0.36	0.84	9 10 6	4 6 3 7 5 8	82327	11 0 0 1.92	0.99 3 12 2 9 5 10 5 5 3	41019	2 2 0

segregating in the family). FBR = female breast cancer under 60 years. MBR = male breast cancer OV = ovarian cancer (2 to BRCA1).

Table 1 also gives the posterior probability of a kindred having a BRCA2 mutation based on LOD scores and prior probability of a recombinant in Kindred 107 with marker tdj3820 at the left boundary, and a second recombinant in Kindred 2043 with marker tdj260. Since the deletion has been driven by BRCA2 itself, this deletion is referred to as the Schutte/Kern deletion in FIG. 1 (Schutte et al., 1995). The S

EXAMPLE 2 Development of Genetic and Physical Resources in the Region of Interest

To increase the number of highly polymorphic loci in the BRCA2 region, we developed a number of STR markers in our lab. STSs in the desired region were used to identify YACs which contained them. These YACs were then used to identify subclones with a number of repeats and/or are of near- perfect fidelity to the pattern. Both of these characteristics are known to increase the polymorphism of STRs. Primers complementary to the end of the repeat. Based on this unique sequence, a primer was made to sequence back across the repeat to confirm their physical localization. New markers which satisfied these criteria were then typed in a set of unrelated individuals.

Using the procedure described above, novel STRs were found from these YACs which were both polymorphic and localized to the BRCA2 region.

EXAMPLE 3 Identification of Candidate cDNA Clones for the BRCA2 Locus by Genomic Analysis of the Contig Region

1. General Methods

Complete screen of the plausible region. The first method to identify candidate cDNAs, although labor intensive, used Inoue's method to identify candidate cDNA clones for future analysis. The clones were screened for putative coding sequences by either of two methods.

The P1 clones to be analyzed were digested with a restriction enzyme to release the human DNA from the vector DNA. The DNA was then digested with Not I restriction enzyme (Boehringer-Mannheim, Cat. #1004760). The eluted Not I digested DNA (~15 kb to 25 kb) was then digested with EcoRI restriction enzyme (Boehringer-Mannheim, Cat. #1004760). The labeled DNA was spermine precipitated (add 100 μ l TE, 5 μ l 0.1 M spermine per the manufacturer's instructions (Gibco/BRL, Cat. #5279SA). The C_0 t-1 blocked probe was incubated on the filters in the presence of a competitor DNA in XAR-5 film with an intensifying screen. Thus, the blots were hybridized with either the pool of Eco-RI fragments from the in-house library or the pool of Eco-RI fragments from the library.

The human DNA from clones in the region was isolated as whole insert or as EcoRI fragments and labeled as described above.

Most of the cDNA libraries used to date in our studies (libraries from normal breast tissue, breast tissue from a woman in her 8th month of pregnancy, and human fetal brain) were grown in C600Hfl bacterial host cells. Normal breast tissue and malignant breast tissue samples were isolated from the same woman. The cDNAs were then cloned into the Lambda Zap II vector, and grown in XL1-blue strain of bacteriophage (Clonetech Cat. HL11810), human placenta (Clonetech Cat 1075b), and human skeletal muscle (Clonetech Cat. HL1124b).

The cDNA libraries were plated with their host cells on NZCYM plates, and filter lifts are made in duplicate from each plate. The filter lifts which correspond to genes located within the candidate cosmid clone. cDNAs identified by this method were picked, re-plated, and analyzed by Southern blot analysis and sequencing. Clones were either purified as plasmid through in vivo excision of the plasmid from the Lambda Zap II vector.

The Southern blot analysis was performed in duplicate, one using the original genomic insert DNA as a probe to verify that the clones are unique were sequenced and the DNA analyzed to determine if the sequences represent known or unique genes. All cDNAs were also analyzed by Northern blot analysis to verify their location. To map the extent of the cDNA clones to the BRCA2 region, tumor RNAs. They are also analyzed by PCR on clones in the BRCA2 region to verify their location. To map the extent of the cDNA clones to the BRCA2 region, analysis.

We have screened the normal breast, 8 month pregnant breast and fetal brain cDNA libraries with Eco RI fragments from the in-house library.

Analysis of hybrid-selected cDNA. cDNA fragments obtained from direct selection were checked by Southern blot hybridization to identify independent clones that overlapped.

The direct selection of cDNA method (Lovett et al., 1991; Futreal, 1993) is utilized with P1 and BAC DNA as the probe. The P1 and BAC DNA primers. Target cDNA is generated from mRNA derived from tissue samples, e.g., breast tissue, by synthesis of either random or specific primers. Sequences are denatured and mixed with human C₀t-1 DNA to block repetitive sequences. Solution hybridization is carried out. Selected cDNA is subjected to further rounds of enrichment before cloning into a plasmid vector for analysis.

HTF island analysis. A method for identifying cosmids to use as probes on the cDNA libraries was HTF island analysis. HTF islands are 100-200 bp long DNA sequences with a high frequency of C₀t-1 dinucleotides. Enzymes known to be useful in HTF-island analysis are Ascl, NotI, BssHII, EagI, SacII, NaeI, NarI, SmaI, and XbaI.

Analysis of candidate clones. One or more of the candidate genes generated from above were sequenced and the information used to determine amino acid sequences. This was accomplished using Genetic Data Environment (GDE) version 2.2 software and the Basic Local Alignment Search Tool (BLAST) workstations. Sequences reconstructed from collections of cDNA clones identified with the cosmids and P1s have been generated.

Mutation screening. To screen for mutations in the affected pedigrees, two different approaches were followed. First, genomic DNA was amplified. An amplified fragment will be larger than predicted from the cDNA sequence or will not be present in the amplified mixture. By comparing the results to the kindreds.

A second approach that is much more rapid if the intron/exon structure of the candidate gene is complex involves sequencing of genomic DNA. Significant extent in lymphocytes, such experiments usually produce amplified fragments that can be sequenced directly with the candidate gene.

The products of such sequencing reactions were analyzed by gel electrophoresis to determine positions in the sequence that correspond to the kindreds.

Any sequence within the BRCA2 region that is expressed in breast is considered to be a candidate gene for BRCA2. Comparison of the sequence to the BRCA2 gene.

2. Specific Methods

Hybrid selection. Two distinct methods of hybrid selection were used in this work.

Method 1: cDNA preparation and selection. Randomly primed cDNA was prepared from poly (A)⁺ RNA of mammary gland, and BAC DNA as described previously. (Parimoo et al., 1991; Rommens et al., 1994). Groups of two to four overlapping P1 and/or BAC DNA fragments were digested with EcoRI, separated on agarose gels, and ligated into pBluescript (Stratagene) that had been digested with EcoRI and treated with calf alkaline phosphatase.

Characterization of Retrieved cDNAs. 200 to 300 individual colonies from each ligation (from each 250 kbases of genomic DNA) were picked and subsequently lysed with standard procedures. Initial analysis of the cDNA clones involved a prescreen for ribosomal sequences.

Approximately 10-25% of the clones were eliminated as they hybridized strongly with radiolabeled cDNA obtained from total RNA. The remaining clones by hybridization to restriction digests of DNAs of the starting clones, of a hamster hybrid cell line (GM10898A) that contained the starting clones, approximately 85% mapped appropriately to the starting clones.

The BRCA2 gene structure was determined by comparison of cDNA and genomic sequences. BRCA2 is composed of 27 exons. BRCA2 appears to be unique, with no close homologs in the human genome.

Expression studies of BRCA2. Hybridization of labeled cDNA to human multiple tissue Northern filters revealed an 11-12 kb transcript. PCR experiments using a BRCA2 cDNA amplicon were performed on five breast and three prostate cancer cell line RNAs. BRCA2 was amplified more efficiently from breast than from thymus.

Germline mutations in BRCA2. Individuals from eighteen putative BRCA2 kindreds were screened for BRCA2 germline mutations. A BRCA2-linked BRCA2 haplotype. Each of the 18 kindreds has a posterior probability of harboring a BRCA2 mutation of at least 69%. In 18 kindreds, 18 individuals were screened for mutations in multiple individuals from nine kindreds using either cDNA or genomic DNA (Table 3). Individuals with mutations at exon junctions.

TABLE 3

Effect	UT-107 ¹	20	18	23	5.00
4706 del4	11	1493	termination codon at 1502	UT-2367 ¹	6 5 1 0 2.09 0.99
1.09	1.00	8525 delC	18	2766	termination codon at 2776
2204 ²	3 1 0 4 0.51	0.98	999 del5	9	257
0.39	0.79	NDUT-2263 ²	3 2 0 1 nd 0.9	NDUT-2171 ²	5 4 2 0 nd nd ND

cDNA sample available. IR inferred regulatory mutation nd not determined Ov Ovarian Cancer ND none detected FBC

Sequence alterations were identified in 9 of 18 kindreds. All except one involved nucleotide deletions that altered the reading frame.

A subset of kindreds was tested for transcript loss. cDNA samples were available for a group of nine kindreds, but three of the kindreds had a polymorphic site. One polymorphic site was interpreted as evidence for a mutation leading to reduction in mRNA levels. In only one of the six cases, some mutations in the BRCA2 coding sequence may destabilize the transcript in addition to disrupting the protein sequence.

Role of BRCA2 in Cancer. Most tumor suppressor genes identified to date give rise to protein products that are absent, nonfunctional, or overexpressed (Srivastava et al., 1993). A similar dominant negative mechanism of action has been proposed for some adenomatous polyposis coli (APC). The BRCA2 coding sequence is consistent with production of either dominant negative proteins or nonfunctional proteins.

EXAMPLE 5 Analysis of the BRCA2 Gene

The structure and function of BRCA2 gene are determined according to the following methods.

Biological Studies. Mammalian expression vectors containing BRCA2 cDNA are constructed and transfected into appropriate cells. Phenotypic reversion in cultures (e.g., cell morphology, doubling time, anchorage-independent growth) and in animals (e.g., tumor formation) is monitored.

Molecular Genetics Studies. In vitro mutagenesis is performed to construct deletion mutants and missense mutants (by site-directed mutagenesis).

Mechanism Studies. The ability of BRCA2 protein to bind to known and unknown DNA sequences is examined. Its ability to interact with other proteins is also studied.

Clones with the desired specificities are expanded and grown as ascites in mice or in a hollow fiber system to produce sufficient quantities of antibody.

EXAMPLE 9 Sandwich Assay for BRCA2

Monoclonal antibody is attached to a solid surface such as a plate, tube, bead, or particle. Preferably, the antibody is attached to a solid surface at room temperature. Next the sample fluid is decanted, and the solid phase is washed with buffer to remove unbound material. The solid phase with the second antibody is incubated for two hrs at room temperature. The second antibody is decanted and the solid phase is washed with buffer.

The amount of bound label, which is proportional to the amount of BRCA2 peptide/protein present in the sample, is quantitated by scintillation counter.

EXAMPLE 10 The 6174delT Mutation is Common in Ashkenazi Jewish Women Affected by Breast Cancer

The 6174delT mutation (see Table 3) has been found to be present in many cases of Ashkenazi Jewish women who have been identified as probands affected with breast cancer on or before 41 years of age with or without a family history of breast cancer. Inclusion criteria for the study was a proband was affected with breast cancer between the ages of 41 and 51 with two or more second degree relatives affected with breast cancer. The study was conducted at several genetic counseling clinics, with an effort to offer study participation to all eligible patients. Family history was obtained by a self-reporting questionnaire.

Mutation Detection

The BRCA2 6174delT mutation was detected by amplifying genomic DNA from each patient according to standard polymerase chain reaction (PCR) protocols.

BC11-RP: GGGAAGCTTCATAAGTCAGTC (SEQ ID NO: 115) (forward primer) and

BC11-LP: TTTGTAATGAAGCATCTGATACC (SEQ ID NO: 116) (reverse primer).

The reactions were performed in a total volume of 10.0 μ l containing 20 ng DNA with annealing at 55° C. This produces a 1 bp deletion in the 6174delT mutation. The gels were then dried and autoradiographed. All the cases exhibiting the 1 bp deletion were sequenced to confirm the 6174delT mutation. The noncoding strand was sequenced. For one set the PCR primers were:

TD-SFB: AATGATGAATGTAGCACGC (SEQ ID NO: 117) (forward primer) and

CGORF-RH: GTCTGAATGTTTCGTTACT (SEQ ID NO: 118) (reverse primer).

This results in an amplified product of 342 bp in wild-type and 341 bp for samples containing the 6174delT mutation. For the other set of primers, the product is 183 bp in a fragment of 183 bp in wild-type samples and 182 bp in samples containing the 6174delT mutation. This was sequenced to confirm the 6174delT mutation.

Results

Six out of eighty women of Ashkenazi Jewish ancestry with breast cancer before the age of 42 had the 6174delT mutation. Four of the six cases with the 6174delT mutation had a family history of breast or ovarian cancer in a first or second degree relative.

cancer at age 42-50 and a history of at least one additional relative affected with breast or ovarian cancer provided an additional estimate of the carrier frequency of the 6174delT mutation in individuals of Ashkenazi Jewish ancestry will emerge. From the data presented, and assuming a penetrance similar to BRCA1 mutations (Offit et al., 1996; Langston et al., 1996), a precise estimate of the carrier frequency of the 6174delT mutation in individuals of Ashkenazi Jewish ancestry will emerge.

TABLE 4					Number of subjects	Number with Group						
Jewish ^a	Before age 37	40	4	(10)	age 37-41	40	2	(5)	Group 2	Diagnosis ages 42-50	27	2

diagnosed with breast or ovarian cancer, one before age 50.

EXAMPLE 11 BRCA2 Shows a Low Somatic Mutation Rate in Breast Carcinoma and Other Cancers Including Ovarian and Pancreatic

BRCA2 is a tumor suppressor gene. A homozygous deletion of this gene may lead to breast cancer as well as other cancers (e.g., ovarian cancer, pancreatic cancer, and prostate cancer). Heterozygosity or LOH. Mutations in both alleles may also lead to development of cancer. For studies here, an analysis of somatic mutations in BRCA2. Since compound mutant heterozygotes and mutant homozygotes are rare, tumor suppressor gene inactivation is assumed to be the cause of cancer.

Identification of tumors and cell lines that exhibit LOH

A group of 104 primary breast tumor samples and a set of 269 cell lines was tested for LOH in the BRCA2 region. For primary tumors, the following STRs were used:

(1) mM4247.4A.2F1 ACCATCAAACACATCATCC (SEQ ID NO: 119)

mM4247.4A.2R2 AGAAAGTAACTTGGAGGGAG (SEQ ID NO: 120)

(2) STR257-FC CTCCTGAAACTGTTCCCTTGG (SEQ ID NO: 121)

STR257-RD TAATGGTGCTGGGATATTTGG (SEQ ID NO: 122)

(3) mMB561A-3.1FA2 GAATGTCGAAGAGCTTGTC (SEQ ID NO: 123)

mMB561A-3.1RB AACATACGCTTAGCCAGAC (SEQ ID NO: 124)

The PCR products were resolved using an ABI 377 sequencer and quantified with Genescan software (ABI). For tumors, the following STRs were used. The tumor miscalled based on later analysis of single base polymorphisms. The heterozygosity indices for the markers are: STR257-FC (0.99), STR257-RD (0.99), and mMB561A-3.1FA2 (0.99). The heterozygosity index for an individual (assuming linkage equilibrium) is only one in 250. Due to the presence of normal cells in the primary tumor samples, the heterozygosity indices for the markers are: STR257-FC (0.99), STR257-RD (0.99), and mMB561A-3.1FA2 (0.99). heights from the tumor (FIGS. 5A-5D). Based on this analysis, 30 tumors (29%) were classified as having LOH at the BRCA2 region.

LOH was assessed in the set of cell lines in a different fashion. Since homozygosity of all three STRs was improbable, and the heterozygosity index for an individual (assuming linkage equilibrium) is only one in 250, the heterozygosity index for a particular tumor cell type under consideration. For example, 4/6 ovarian cell lines and 31/62 lung cancer lines displayed LOH at the BRCA2 region.

Sequence Analysis of LOH Primary Breast Tumors and Cell Lines

The 30 primary breast cancers identified above which showed LOH in the BRCA2 region were screened by DNA sequencing. In a radioactive mutation screen, the amplified products were purified by Qiagen beads (Qiagen, Inc.). DNA sequence was generated by electrophoresis on ABI 377 sequencers. Samples were gridded into 96-well trays to facilitate PCR and sequencing. Dropouts were confirmed by sequencing a newly amplified PCR product to exclude the possibility that the sequence alteration was due to

TABLE 5

Type	# LOH/# Screened	Percentage LOH
Lung	31/62	50%
Lymphoma	0/4	0%
Melanoma	17/81	21%
Neuroblastoma	9/42	21%

LOH analysis of cell lines and primary breast tumors. Percentage LOH was calculated two ways: as total and as a mean of

Of the 30 samples, two specimens contained frameshift mutations, one a nonsense mutation, and two contained missense mutations. Sequence variants were also present in the corresponding normal DNA from these cancer patients. To exclude the unlikely possibility that the mutations were germline, three of these revealed missense changes that were also found in the normal samples. Thus, in a set of 42 breast carcinoma patients. The missense variants are rare; they were each observed only once during analysis of 115 chromosomes. From the

Of the 85 cell lines which displayed LOH (see Table 5), 58 were also screened for sequence changes. Greater than 95% of the BT111 primary tumor sample and to a previously detected germline frameshift (Tavtigian et al., 1996). This suggests that the

Detection of a probable germline BRCA2 mutation in a pancreatic tumor cell line suggests that BRCA2 mutations may precede the formation of a pancreatic xenograft (Schutte et al., 1995). Because only three pancreatic cell lines were examined in our study, further investigation

TABLE 6A

Sample	Type	LOH	Change	Effect	Germline			
Frameshift	yes	4F8	Ovarian	yes	C2117T Thr→Ile BT163	Primary breast	no	A2411C
Thr→Met BT111	Primary breast	yes	6174delT	Frameshift	yes	4G3	Panc	
Gln→His	yes	3D5	Melanoma	yes	A9537G Ile→Met BT85	Primary breast	yes	A10204

Germline mutations identified in BRCA2. Listed are the mutation positions based on the Genbank entry of BRCA2 (Schutte et al., 1995).

TABLE 6B

Position	Change	Effect	Frequency
A/G	silent	0.35 PM(3668) A/G	Asn→Ser
0 (0.15)	PM(4035) T/C	silent	0.24 (0.10) PM(74)
silent	<0.01		

Common polymorphisms and silent substitutions detected in BRCA2 by DNA sequencing. Since some rare silent variants were not shown in parentheses (Tavtigian et al., 1996). Numbering is as in Table 6A.

Industrial Utility

As previously described above, the present invention provides materials and methods for use in testing BRCA2 alleles of a

protective effect of an early, full term pregnancy. Therefore, women at risk could consider early childbearing or a therapy that could be highly motivated to have regular mammograms, perhaps starting at an earlier age than the general population. Ovarian precursor lesions. With the evolution of the method and the accumulation of information about BRCA2 and other causative

Women with breast cancers may follow different surgical procedures if they are predisposed, and therefore likely to have a product. Alternatively, the therapeutic agent could be another molecule that mimics the deleterious gene's function, either a counteract the effect of the deleterious allele. These gene therapies may take many forms and may be directed either toward

It will be appreciated that the methods and compositions of the instant invention can be incorporated in the form of a variety of be construed as restrictive.

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List of Patents and Patent Applications

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U.S. Pat. No. 3,850,752

U.S. Pat. No. 3,939,350

U.S. Pat. No. 3,996,345

U.S. Pat. No. 4,275,149

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U.S. Pat. No. 4,868,105

U.S. Pat. No. 5,252,479

EPO Publication No. 225,807

European Patent Application Publication No. 0332435

Geysen, H., PCT published application WO 84/03564, published 13 Sep. 1984

Hitzeman et al., EP 73,675A

PCT published application WO 93/07282

SEQUENCE LISTING(1) GEN

TOPOLOGY: linear(ii) MOLECULE TYPE: cDNA(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) ORIGINAL SOURCE:
NO:1:GGTGGCGCGAGCTTCTGAAACTAGGCGGCAGAGGCGGAGCCGCTGTGGCACTGCTGCGCC60TCTGCTGCGC
ATATCGTAGGTAAAAATGCCTATT237MetProlleGGATCCAAAGAGAGGCCAACATTTTTTGAATTTTTAAGACACGCTG
ACCTATAATTCTGAACCTGCAGAAGAATCTGAA381SerSerGluAlaProProTyrAsnSerGluProAlaGluGluSerGlu404550CA

GlnLeuAlaSerThrProllellePheLysGlu707580CAAGGGCTGACTCTGCCGCTGTACCAATCTCCTGTAAAAGAATTAGAT52
GTGAAAACATAAAATGGATCAAGCAGATGATGTTTCC621SerLeuArgThrValLysThrLysMetAspGlnAlaAspAspValSer12012
hrHisValThrProGlnArgAspLysSerValValCysGlySer150155160TTGTTTCATACACCAAAGTTTGTGAAGGGTCGTCAGACA
AGTTCTTTAGCTACACCACCCACCCTTAGTTCTACTGTGCTCATAGTC861SerSerLeuAlaThrProProThrLeuSerSerThrVa
AAT957AsnValLysSerTyrPheSerAsnHisAspGluSerLeuLysLysAsn230235240GATAGATTTATCGCTTCTGTGACAGACAG
ys260265270275GTAAATAGCTGCAAAGACCACATTGGAAAGTCAATGCCAAATGTCCTA1101ValAsnSerCysLysAspHis
AAAATCTACAAAAGTA1197PheSerLeuCysPheSerLysCysArgThrLysAsnLeuGlnLysVal310315320AGAACTAGCAAGAC
LysGluLysTyrSerPheVal340345350355TCTGAAGTGGAAACCAATGATACTGATCCATTAGATTCAAATGTAGCA1341Ser
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amino acids(B) TYPE: amino acid(D) TOPOLOGY: linear(ii) MOLECULE TYPE: protein(xi) SEQUENCE DESCRIPTION: S
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DESCRIPTION: SEQ ID NO:16:GTGCTCATAGTCAGAAATGAAG22(2) INFORMATION FOR SEQ ID NO:17:(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 19 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:TCTTCCCATCCTCACAGTAA21(2) INFORMATION FOR SEQ ID NO:22:(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 21 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:GTAGGAAAATGTTTCATTTAA21(2) INFORMATION FOR SEQ ID NO:23:(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 20 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:ATCTAAAGTAGTATTCACA21(2) INFORMATION FOR SEQ ID NO:28:(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 20 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:ATTGTCAGTTACTAACACAC20(2) INFORMATION FOR SEQ ID NO:29:(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 20 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:GTGTCATGTAATCAAATAGT20(2) INFORMATION FOR SEQ ID NO:30:(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 19 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:CAGGTTTAA21(2) INFORMATION FOR SEQ ID NO:35:(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 22 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:AACAGTTGTAGATACCTCTGAA22(2) INFORMATION FOR SEQ ID NO:36:(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 22 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:GACTTTTTGATACCCTGAAATG22(2) INFORMATION FOR SEQ ID NO:41:(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 22 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:TTTAGTGAATGTGATTGATGGT22(2) INFORMATION FOR SEQ ID NO:42:(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 21 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:AGAACCAACTTTGTCCTTAA20(2) INFORMATION FOR SEQ ID NO:47:(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 19 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:47:GCTACCTCCAAAAGTGA19(2) INFORMATION FOR SEQ ID NO:48:(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 22 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:48:GTGTAAAGCAGCATATAAAAAT22(2) INFORMATION FOR SEQ ID NO:49:(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 21 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:49:CTTAAATGTTTAA21(2)

DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(x) single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) OR pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: DNA (genomic)(iii) SEQUENCE CHARACTERISTICS:(A) LENGTH: 21 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TO NO:53:GGCTTTTATTCTGCTCATGGC21(2) INFORMATION FOR SEQ ID NO:54:(i) SEQUENCE CHARACTERISTICS:(A) L Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:54:CCTCTGCAGAAGTTTCTCAC21(2) INFORMATION FOR SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:55:AA DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(x) single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) OR base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: DNA (genom SEQUENCE CHARACTERISTICS:(A) LENGTH: 19 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TO NO:59:GCCTTAGCTTTTTACACAA19(2) INFORMATION FOR SEQ ID NO:60:(i) SEQUENCE CHARACTERISTICS:(A) L sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:60:TTTTTGATTATATCTCGTTG20(2) INFORMATION FOR SEQ ID NO:61:TTATTCTC (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(xi) SE single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) OR base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: DNA (genom SEQUENCE CHARACTERISTICS:(A) LENGTH: 21 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TO NO:65:GAAGATAGTACCAAGCAAGTC21(2) INFORMATION FOR SEQ ID NO:66:(i) SEQUENCE CHARACTERISTICS:(A) Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:66:TGAGACTTTGGTTCCTAATAC21(2) INFORMATION FOR SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:67:AG TYPE: DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sa STRANDEDNESS: single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI LENGTH: 21 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: ID NO:71:(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 21 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: NO:71:ACTCTTTCAAACATTAGGTCA21(2) INFORMATION FOR SEQ ID NO:72:(i) SEQUENCE CHARACTERISTICS:(A) Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:72:TTGGAGAGGCAGGTGGAT18(2) INFORMATION FOR S SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:73:CT DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(x) single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) OR pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: DNA (genomic)(iii) SEQUENCE CHARACTERISTICS:(A) LENGTH: 21 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TO NO:77:GAATACAAAACAGTTACCAGA21(2) INFORMATION FOR SEQ ID NO:78:(i) SEQUENCE CHARACTERISTICS:(A) Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:78:CACCACCAAAGGGGAAA18(2) INFORMATION FOR S SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:79:AA DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(x) single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) OR pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear(ii) MOLECULE TYPE: DNA (genomic)(iii) SEQUENCE CHARACTERISTICS:(A) LENGTH: 22 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TO

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NO:95:TGAATGTTATATATGTGACTTTT23(2) INFORMATION FOR SEQ ID NO:96:(i) SEQUENCE CHARACTERISTICS:
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SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:97:CC
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NO:101:TTTGTTGTATTTGTCCTGTTTA22(2) INFORMATION FOR SEQ ID NO:102:(i) SEQUENCE CHARACTERISTICS:
Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:102:ATTTTGTAGTAAGGTCATTTTT23(2) INFORMATION F
ANTI-SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1
MOLECULE TYPE: DNA (genomic)(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANIS
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Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:108:CTATTTTGATTTGCTTTTATTATT24(2) INFORMATION
ANTI-SENSE: NO(vi) ORIGINAL SOURCE:(A) ORGANISM: Homo sapiens(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1
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TYPE: other nucleic acid(A) DESCRIPTION: /desc = "Primer"(iii) HYPOTHETICAL: NO(iv) ANTI-SENSE: NO(xi) SEQUEN
TOPOLOGY: linear(ii) MOLECULE TYPE: other nucleic acid(A) DESCRIPTION: /desc = "Primer"(iii) HYPOTHETICAL: NO

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25		Wooster, R. et al. (1995). " Identification of the breast cancer susceptibility gene BRCA2 ",
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* Cited by examiner

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US7897356	Nov 12, 2009	Mar 1, 2011	Caris Life Sciences	Methods
US7933722	May 10, 2006	Apr 26, 2011	Synergz Bioscience Limited	Methods
US8076065	May 19, 2006	Dec 13, 2011	Synergz Bioscience Limited	Methods
US8700335	May 18, 2007	Apr 15, 2014	Caris Mpi, Inc.	System a
US8768629	Feb 12, 2010	Jul 1, 2014	Caris Mpi, Inc.	Molecula

CLASSIFICATIONS

U.S. Classification	435/69.1 , 435/320.1 , 435/375 , 530/828
International Classification	A61K48/00 , C07K14/47 , C12N15/12
Cooperative Classification	Y10S530/828 , A61K48/00 , C07K14/4703
European Classification	C07K14/47A1A

LEGAL EVENTS

Date	Code	Event	
Apr 22, 2010	FPAY	Fee payment	Year of fee payment: 12
Apr 28, 2006	FPAY	Fee payment	Year of fee payment: 8
Apr 27, 2006	AS	Assignment	Owner name: UTAH RESEARCH FOUNDATION, UN Free format text: ASSIGNMENT OF ASSIGNORS INT Effective date: 20060420
May 17, 2002	FPAY	Fee payment	Year of fee payment: 4
Oct 9, 1996	AS	Assignment	Owner name: ENDO RECHERCHE INC., CANADA Free format text: ASSIGNMENT OF ASSIGNORS INT Effective date: 19960819 Owner name: HOSPITAL FOR SICK CHILDREN, CAN Free format text: ASSIGNMENT OF ASSIGNORS INT Owner name: HSC RESEARCH & DEVELOPMENT L Free format text: ASSIGNMENT OF ASSIGNORS INT

Effective date: 19960617

Owner name: MYRIAD GENETICS, INC., UTAH

Free format text: ASSIGNMENT OF ASSIGNORS INT

Owner name: PENNSYLVANIA, THE TRUSTEES OF

Free format text: ASSIGNMENT OF ASSIGNORS INT

Effective date: 19961002

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